



SION (WEST), MUMBAI- 400022

FACULTY: Science

PROGRAM: MSc Part II

Subject: Biotechnology

Academic year: 2024-2025

Semester III and IV

Credit-based semester and grading syllabi approved by the Board of Studies in Biotechnology to be brought into effect from June 2024.

PREAMBLE

Biotechnology, broadly defined, includes any technique that uses living organisms, or parts of such organisms, to make or modify products, to improve plants or animals, or to develop microorganisms for specific use. The interdisciplinary nature of biotechnology integrates living systems including animal, plant and microbes and their studies from molecular biology to cell biology, from biochemistry to biophysics, from genetic engineering to stem cell research, from bioinformatics to genomics-proteomics, from environmental biology to biodiversity, from microbiology to bioprocess engineering, from bioremediation to material transformation and so on.

Biotechnology is the science of today and tomorrow. It has applications in all major service sectors. i.e. health, agriculture, industry, environment etc. Biotechnology as an application science has taken firm footing in many countries, abroad where a number of transgenic crops, genetically modified food and recombinant therapeutic molecules for human and animal health are available in the market. Biotechnology as a science of service to human society is yet to make inroads in India

With the advent of the World Wide Web in the early nineties and its subsequent growth, the latest research trends have become accessible from drawing rooms across the globe. This acted as a positive feedback mechanism in increasing the pace of research in all fields including Chemical Engineering and Bio-technology. This was the motivation for an in depth analysis of what is actually required for today's technology. It is also important to take advantage of the freely available software to enhance the quality and quantity of material that can be covered in the classroom.

This restructured syllabus is therefore intended to combine the principles of physical, chemical and biological sciences along with developing advanced technology. The postgraduate curriculum is prepared to impart primarily basic knowledge of the respective subject from all possible aspects. In addition, students will be trained to apply this knowledge particularly in day-to-day applications of biotechnology and hence get a flavor of research

PROGRAM OUTCOMES

The expected graduate attributes are directed towards the following:

- Applying the knowledge of various courses learned under the program to break down complex problems to simple components by designing processes for problem solving
- Utilizing the acquired contextual knowledge in an interdisciplinary framework. Integrating research-based knowledge and research-based methods involving problem definition, analysis and interpretation of data followed by its consolidation to arrive at valid conclusions
- Facilitating to write and document effectively; make crisp presentations and reports and convey the message/ instructions/findings clearly
- Equipping to select, create and apply the appropriate tools and techniques through electronic media for the purpose of understanding and analyzing data and drawing inference keeping in mind its limitations and disadvantages
- Understanding the need for sustainable development and concern for environmental issues
- Applying the acquired contextual knowledge in assessing public health and safety; addressing gender, ethnic and environmental issues in addition to performing with decisive responsibility.

PROGRAM SPECIFIC OUTCOMES

The program has been designed to expose the students to the latest developments in the areas of diagnostics, therapeutic techniques and instrumentation. This program is aimed at empowering students for a career in research and also to provide trained manpower for the fast-growing Biotech companies.

A Postgraduate student upon completion of this program is expected to gain the following attributes:

- Competence for research and innovation in the field of Biotechnology
- Design and execute experiments applying the concepts learnt and thereby being able to translate theoretical knowledge to practical knowledge
- Prepare, plan and execute a research project independently.
- Critically evaluate and interpret results

Semester III

Course Category	Course code	Course Title	Credits	Lectures (Hrs.)/week
Core Course 1	SIPBTCC611	PTC and ATC	4	4
Core Course 2	SIPBTCC612	Medical Biotechnology	4	4
Core Course 1	SIPBTCCP611	Practical	2	4
Core Course 2	SIPBTCCP612	Practical	2	4
Discipline-specific Elective	SIPBTEL611	Biostatistics	3	3
Discipline-specific Elective	SIPBTELP611	Practical	1	2
RP	SIPBTCRP611	Research Proposal	6	
			Total Credits 22	

Semester IV

Course Type	Course code	Course Title	Credits	Lectures (Hrs.)/week
Core Course 1	SIPBTCC621	Omics and Bioinformatics	4	4
Core course 2	SIPBTCC622	Nanotechnology and Clinical Studies	4	4
Core course 1	SIPBTCCP621	Practical	2	4
Core course 2	SIPBTCCP622	Practical	2	4
Discipline-specific Elective	SIPBTEL621	Biosafety	3	3
Discipline-specific Elective	SIPBTELP621	Practical	1	2
RP	SIPBTCRP621	Research Project	6	
			Total Credits 22	

Draft syllabus (MSc Part II)

Semester III

Course Type	Course Title	Course description	Credits	Lectures (Hrs.)/ week
Core Course 1	PTC and ATC	<p>Unit I -</p> <p>Biosynthesis- batch, continuous cultures, immobilized plant cells, Biotransformation of precursors by cell culturing, metabolic engineering for the production of secondary metabolites, Hairy root culture, elicitation, synthetic seeds. Genetic markers in plant breeding-- Classical markers, DNA markers (RFLP, RAPD, AFLP, SSR, SNP)</p> <p>Unit II-</p> <p>Cryopreservation - Principle and types. Germplasm conservation, Transgenic plants -Edible vaccine, Golden rice Virus, herbicide, fungus and bacteria resistant plants, Genetic manipulations for fruit ripening, flower wilting & flower pigmentation. Plants as bioreactors</p> <p>Unit III-</p> <p>Biology of cultured cells – cell adhesion, cell proliferation, Differentiation, Cell signaling and senescence, Culture vessels, substrates and specialized systems, Culture Media and supplements, Serum-free media, Primary culture: Types, isolation of tissues, culturing of different cells, Microbial contamination and Cross contamination.</p>	4	4

		<p>Unit IV-</p> <p>Cell lines: cell line designation, choosing a cell line, routine maintenance, replacement of media, Subculture and propagation, Split ratio, Organotypic culture, Transformation & immortalization of cell line, Cytotoxicity & Quantitation.</p>		
Core Course 2	Medical Biotechnology	<p>Unit I -</p> <p>Molecular Diagnostic methods: Fluorescent in situ hybridization (FISH), MFISH, chromosome painting, CGH, Microarray. Target amplification methods- PCR and RT PCR, Isothermal methods- NASBA, TMA, Strand displacement amplification, LAMP. Probe amplification reaction – LCR. Signal amplification method- bDNA assay</p> <p>Unit II-</p> <p>Pathogenesis (Mechanism of infection) and Acute Clinical manifestations (Signs and symptoms) laboratory diagnosis & treatment of Pneumonia, Tuberculosis, MOTT, <i>Helicobacter pylori</i>, Nosocomial (<i>Pseudomonas</i>)</p> <p>Biofilms in medicine: Structure of Biofilm – Extracellular polymeric substances, Biofilm architecture, Stages in formation of Biofilm, Microbial interactions in Biofilms (Quorum sensing), Need for formation of Biofilms by microorganisms, Microorganisms commonly associated with biofilms on indwelling medical devices, Response of biofilms to host defense mechanisms & antimicrobial agents and Recent advances in biofilm management.</p>	4	4

		<p>Unit III-</p> <p>Concept of Host, Reservoir, Source of infection, Carrier, Epidemic, Endemic, Pandemic, Outbreak, importance of epidemiology, Measures of disease frequency – Concept of incidence, prevalence, Incidence rate, cumulative incidence, case fatality, Epidemiological case studies</p> <p>Structure of the virus-coat and envelope protein, genome composition Pathogenesis (Mechanism of infection) and Acute Clinical manifestations (Signs and symptoms), laboratory diagnosis & treatment of HIV, Hepatitis (A,B,C), Influenza (HINI, H5N5), Ebola, HSV, HPV Corona virus (Covid 19) and zoonotic infections.</p> <p>Unit IV-</p> <p>Antigenic structure, virulence factors, source of infection, Transmission, Pathogenesis, Life-cycle, Clinical manifestations, Laboratory diagnosis, Treatment, Prophylaxis, Current research and developments for:</p> <p>Fungal infections- Candidiasis, Tinea</p> <p>Protozoan infections -Malaria, Toxoplasmosis, Amoebiasis, Trypanosomal infection</p> <p>Chlamydia and Rickettsial infections</p>		

Core Course 1	Practical	<p>PTC and ATC</p> <p>1. PTC</p> <p>a. Media preparation</p> <p>b. Seed sterilization</p> <p>c. Callus induction</p> <p>d. Isolation and induction of anther cultures using Hibiscus flowers.</p> <p>e. Protoplast isolation</p> <p>f. Somatic embryogenesis</p> <p>2. ATC</p> <p>a. Trypsinization and staining of animal cells</p> <p>b. Monolayer formation (fibroblast)</p> <p>c. To assay the radical scavenging activity of a tissue hydrolysate – DPPH method</p> <p>d. Techniques of cell preservation</p>	2	4
Core Course 2	Practical	<p>Medical Biotechnology</p> <p>Medical diagnostic – Identification of organisms from specimens (Multidrug resistant <i>S. aureus</i>, <i>Pseudomonas</i> spp., <i>Klebsiella pneumoniae</i>, <i>E. coli</i>)</p> <p>Qualitative and quantitative estimation of biofilm production by microorganisms.</p>	2	4

<p>Discipline Specific Elective</p>	<p>Biostatistics</p>	<p>Unit I -</p> <p>Statistical population, Types of data, Census survey, Sample survey, Random sample. Central Tendency: Mean, Median and Mode. Absolute measures of dispersion: Range, Quartile deviation, Mean deviation, Standard deviation. Relative measures dispersion. Coefficient of skewness and kurtosis based on moments, Karl Pearson and Bowley's measure of skewness</p> <p>Correlation: Definition, Scatter plot, Relationship with covariance, Karl Pearson's correlation coefficient, Simple linear regression, Coefficient of determination (R^2), Concept of multiple correlation and regression.</p> <p>Unit II-</p> <p>Probability distribution: Discrete (e.g., Binomial) and Continuous (e.g., Normal)</p> <p>Sampling distribution: Definition, Sampling from normal and non- normal population, Concept of standard error, The Central Limit Theorem</p> <p>Concept of estimation & confidence limits</p> <p>Hypothesis testing: Null and alternative hypothesis, test of a statistical hypothesis, Type I and Type II errors, Power of a test, level of significance of a test. Concept of p-value.</p> <p>Large sample tests for the significance of: i) single population – a) mean and b) proportion ii) difference in two population a) means and b) proportions</p> <p>t-test for the significance of: i) single population mean ii) difference in two population means– independent and dependent samples. One way analysis of variance</p>	<p>3</p>	<p>3</p>
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		<p>Unit III</p> <p>Non-parametric tests: Definition, Merits, Demerits, Characteristics of Non-parametric Tests, Comparable tests for the parametric tests, Single sample: Sign test, Wilcoxon test, Run test for randomness Two sample: Sign test, Wilcoxon test, Mann-Whitney test, k sample: Kruskal-Wallis test, Friedman Test.</p>		
Discipline Specific Elective	Practical	<ol style="list-style-type: none"> 1. Use of MS Excel for basic calculations 2. Graphical representation using Excel 3. Correlation and regression analysis 4. Finding p-value for a statistical test 5. Research paper presentation with statistical analysis 	1	2

Draft syllabus (MSc Part II)

Semester IV

Course Type	Course Title	Course description	Credits	Lectures (Hrs.)/ week
Core Course 1	Omics and Bioinformatics	<p>Unit I -</p> <p>Introduction to Omics, Genomics and epigenomics; Transcriptomics; Structural genomics; Functional genomics. Metagenomics: concept, strategies. Large scale genome sequencing strategies, Genome databases, Human genome project- goals, conclusions and application.</p> <p>Tools and techniques: NGS, Microarrays, SAGE, RAGE, ChIPseq, qPCR, RNA sequencing, ENCODE. Epigenomic analyses and cancer/ diseases. Bisulfite sequencing Applications in environmental biotechnology, agriculture and health.</p> <p>Unit II-</p> <p>Proteomics and Interactomics -protein- protein interaction and identification of interactions by various methods. Yeast two hybrid systems; Synthetic genetic array (SGA) analysis; Protein microarrays; Reverse phase protein array; NAPPA; Halotag array; SPR; label free assays; biolayer interference; PISA. Application of Proteomics and Genomics in human diseases – screening, testing and treatment of diseases, personalized medicine.</p> <p>Unit III- Biological databases and its types. Submission of data to databases, Querying in databases – Retrieval of desired data; Types of sequence alignment – BLAST – Nucleotide blast and protein Blast – Applications Multiple sequence alignment – Principle, methods, phylogenetic analysis. Protein sequence analysis – Secondary structure analysis protein motifs, protein families Protein structure analysis and applications: Protein</p>	4	4

		<p>databank, Structural classification of proteins -CATH, Protein, Structure visualization and importance. Introduction to Docking studies.</p> <p>Unit IV-</p> <p>Gene Prediction: Categories of Gene Prediction Programs, Gene Prediction in Prokaryotes, Gene Prediction in Eukaryotes, Gene clusters and fusions, DNA motif finding</p> <p>Promoter and Regulatory Element Prediction- Promoter and Regulatory Elements in Prokaryotes, Promoter and Regulatory Elements in Eukaryotes, Prediction Algorithms</p> <p>Designing primers for gene sequence.</p>		
Core Course 2	Nanotechnology and clinical studies	<p>Unit I -</p> <p>Introduction, Examples of nanostructures found in nature, Nanorobotics devices of nature: ATP synthase, motor proteins of cytoskeleton, flagella. Nanometer-scale materials, Quantum dots, Carbon nanotubes Synthesis of nanomaterials: Top-down and bottom-up approaches, Biodirected synthesis, Synthesis using bio-derived templates, Self- assembly of nanoparticles.</p> <p>Unit II-</p> <p>Applications: Nanomedicine, drug delivery, nanosensors, food, cosmetics, agriculture, environment</p> <p>Nanotoxicology: Unique Properties, Factors Responsible for Nanomaterial Toxicity, Mechanisms of Nanoparticle Toxicity, In Vitro Testing Methods for Nanomaterials, Ecotoxicity Analyses of Nanomaterials.</p>	4	4

		<p>Unit III –</p> <p>Pre-clinical toxicology: General principles, systemic toxicology (single dose and repeat dose toxicity studies), Carcinogenicity, Mutagenicity, Teratogenicity, Reproductive toxicity, Local toxicity, Genotoxicity, Animal toxicity requirements.</p> <p>Clinical study reports, Principles in softwares in CDM (Clinical Data Management)</p> <p>Unit IV –</p> <p>Types of clinical trials, single blinding, double blinding, open access, randomized trials and their examples, interventional study.</p> <p>Adverse Drug Reactions (ADR) - ADR classification, Nature and mechanism of ADR, Concept of safety, Phases and types of DATA. The process of Pharmacovigilance - Signal detection, evaluation and investigation, Communication Methods of evaluating effectiveness of action.</p>		
Core Course 1	Practical	<p>Omics and Bioinformatics</p> <ol style="list-style-type: none"> 1. Multiple alignment – Phylogenetic tree 2. BLAST – homologs, orthologs and paralogs 3. Motif finding 4. KEGG 5. Structure of proteins – identification of chains, helices, special groups, metal ions, etc. 6. CATH / SCOP classification of a given protein 7. Signal Peptide detection using TargetP tools 	2	4
Core Course 2	Practical	<p>Nanotechnology Practical</p> <ol style="list-style-type: none"> 1. Nanoparticles synthesis by chemical reduction 	2	4

		<p>2. Nanoparticles synthesis by biological methods</p> <p>3. Spectroscopic analysis of nanoparticles</p>		
Discipline Specific Elective	Biosafety	<p>Unit I -</p> <p>Genetically modified microorganisms, examples and methods, Humulin, Ice minus bacteria, GM bacteria in bioremediation, use of PCR as GMO identification tool, risks and controversies related to use of GMO. Indian GM research information system, About Indian GMO research information system (IGMORIS) Biosafety data of any two approved genes available on the database</p> <p>GE crops – Arabidopsis as a model plant for studies in genetic engineering</p> <p>Unit II-</p> <p>Introduction; Biological Risk Assessment, Hazardous Characteristics of an Agent; Genetically modified agent hazards; cell cultures; Hazardous Characteristics of Laboratory Procedures; Potential Hazards Associated with Work Practices; Safety Equipment and Facility Safeguards; Pathogenic risk and management. Concept of GLP; Guidelines to GLP; Documentation of Laboratory work; Preparation of SOPs; Calibration records; Validation of methods; Documentation of results; Audits & Audit reports</p> <p>Unit III-</p> <p>Microbiological Assays for pharmaceutical products; Regulatory Microbiological testing in pharmaceuticals</p> <p>Concepts on biosafety in Biotechnology; Regulating rDNA technology; Regulating food and food ingredients;</p>	3	3

		Genetically engineered crops, livestock Bioethics; Contemporary issues in Bioethics		
Discipline Specific Elective	Biosafety Practical	Biosafety 1. GMO kit-based validation 2. Biosafety practices in a Biotechnology laboratory 3. Writing SOP for laboratory instruments 4. Validation of HPTLC, GC HPLC, and micropipettes. 5. Sterility testing of injectables	1	3

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